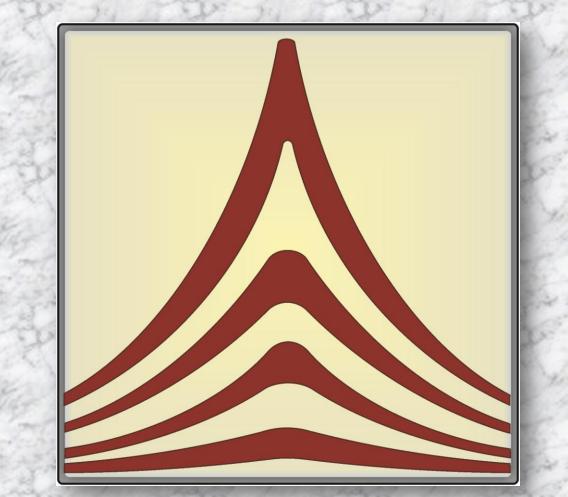


CHARACTERIZATION OF GRAM POSITIVE BACTERIA USING RAMAN SPECTROSCOPY A. Colniță^{1,2}, Nicoleta Elena Dina¹, Dan Vodnar³, Nicolae Leopold¹, Vasile Chis¹, Leontin David¹



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Abstract

In the last years, Raman spectroscopy has become a well-known physicochemical technique for the rapid identification of microorganisms. This powerful analytical method generates a spectroscopic fingerprint from the bacterial sample, which provides quantitative and qualitative information used to characterize, discriminate and identify different types of microorganisms. This work reports the use of Raman and surface-enhanced Raman scattering (SERS) spectroscopic methods for identification, characterization and discrimination of Gram-positive bacterial species. Lactobacillus casei and Listeria monocytogenes are rod shaped, nonsporing, anaerobic bacteria. While L. casei is a well-known probiotic, since it has numerous beneficial effects, L. monocytogenes is the agent of listeriosis, a serious infection caused by eating food contaminated with this type of bacteria. In order to obtain the Raman spectra, the bacteria were harvested and suspended in a small volume of water. The measurements were made on MgF₂ slides using 50x objective. As SERS active substrate we used silver colloids prepared by different reduction methods and the obtained SERS spectra were compared with the Raman spectra. The assignment of the main bands was made by taking into account the position and the relative intensities. We were able to optimize the bacteria sample preparation procedures in order to obtain reproducible SERS spectra that enable us to characterize and discriminate the two Gram-positive species.

Growth conditions - L. casei ATCC 393 routinely cultured overnight at 37 °C in MRS broth (Oxoid, UK). • sample concentration: Sample preparation for Raman / SERS measurements 2 mL bacteria culture were centrifuged (6000 rpm, 10 min) and washed 3 times with deionized water; the harvested pellet was resuspended in 1 mL H_2O for all the measurements.

Growth conditions - L. monocytogenes ATCC 19115 • maintained on Oxford agar (Sifin, Germany) plates at 4°C; a single colony of L. monocytogenes was inoculated into a tube of tryptic soy broth plus 0.7% yeast extract (TSBYE) (Difco Laboratories) and incubated at 35 °C for 24 h.

Sample concentration: 10^{11} CFU/ml. (0.D. 600nm=0.7)

Sample preparation for Raman / SERS measurements

2 mL bacteria culture were centrifuged (6000 rpm, 10 min) and washed 3 times with deionized water; the harvested pellet was resuspended in 250 μ L H₂O for all the measurements.

Experimental methods

Raman / SERS spectra (Department of Biomolecular Physics, Faculty of Physics, Babes-Bolyai University, Cluj-Napoca)

• Renishaw inVia Raman Microscope with He-Ne laser (632.8 nm); objective: 50x; total laser power: 50mW; resolution: 4 cm⁻¹

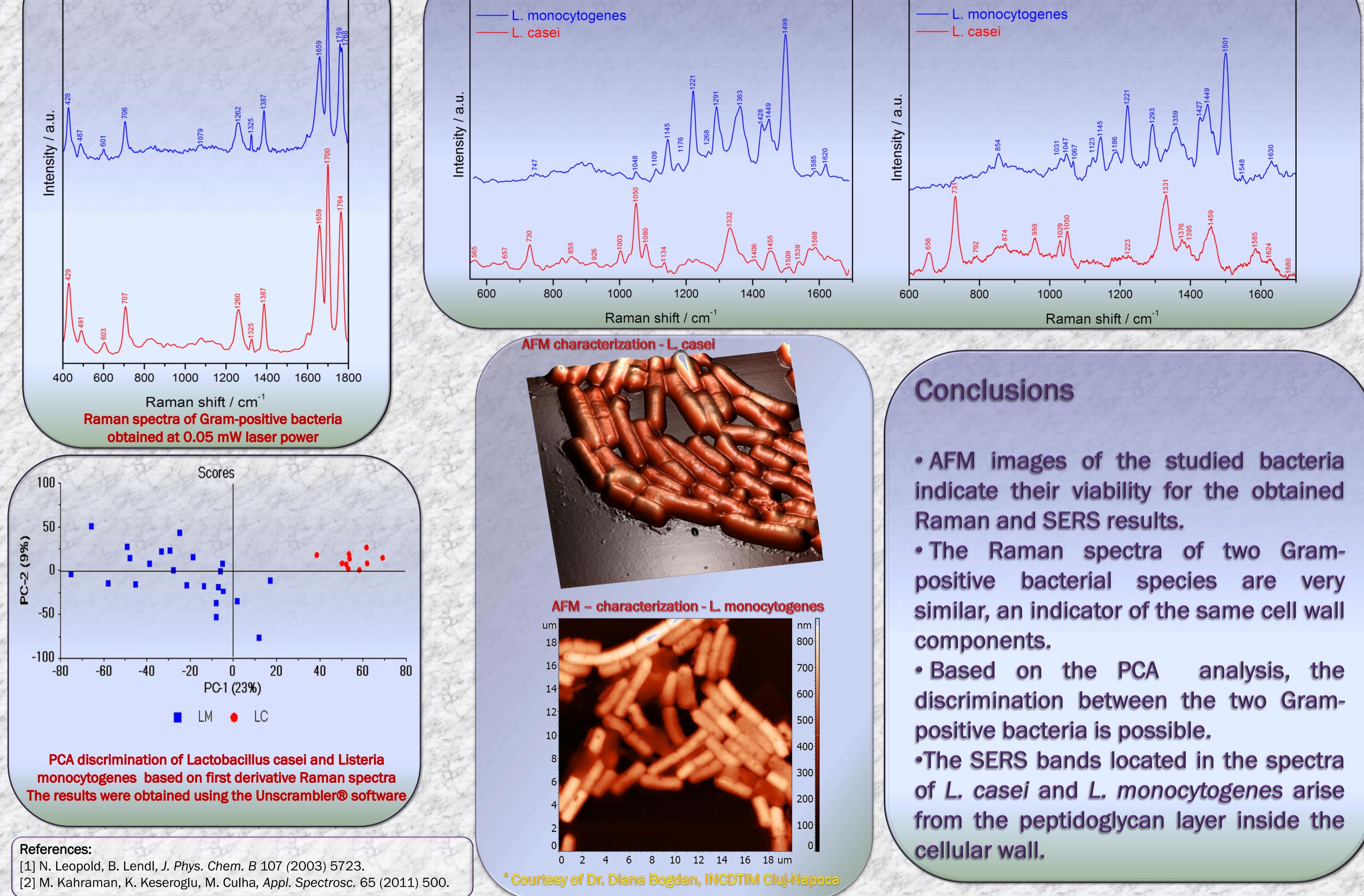
• SERS spectra – AgNPs were prepared by reduction of AgNO₃ with hydroxilamine hidrochloride (hya) [1].

- for in situ detection: to 100 μ I AgNO₃ (10⁻³ M) we added bacteria suspension (5-30 μ I) and 900 μ I hya solution (~10⁻³ M) with basic pH [2].

. monocytogenes casei

SERS - with a priori prepared colloid

SERS – in situ detection



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